## ANTIBIOTIC AC6H, A NEW COMPONENT OF TETROCARCIN GROUP ANTIBIOTICS

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We wish to describe here isolation and structural elucidation of a new antibiotics, AC 6 H , from the culture broth of Micromonospora carbonaceae
subsp. carbonaceae K55-AC6, which was isolated from a soil sample collected at Kawanishi-city, Hyogo Prefecture, Japan.
The strain was inoculated to a seed medium (glucose $1.0 \%$, soybean meal $1.0 \%$, meat extract $0.5 \%$ and $\mathrm{CaCO}_{3} 0.4 \%$ ) in a $500-\mathrm{ml}$ Erlenmeyer flask and incubated on a rotary shaker at $27^{\circ} \mathrm{C}$ for 3 days. The seed culture ( 2 ml ) was transferred to $500-\mathrm{ml}$ Erlenmeyer flasks containing 100 ml of a production medium, consisting of lactose $1.0 \%$, soybean meal $1.0 \%$, meat extract $0.5 \%, \mathrm{CaCO}_{3}$ $0.4 \%$ and Allophosite $1.0 \%$.

After 5 days of cultivation, the culture filtrate ( 5 liters) was passed through a column of Diaion HP-20, which was washed with $60 \%$ aq. MeOH and developed with MeOH ( 2 liters). The active eluate

Table 1. Chemical shift assignments of $\mathrm{AC} 6 \mathrm{H} \cdot \mathrm{Na}$ and $\mathrm{AC} 6 \mathrm{D} \cdot \mathrm{Na}$.

| Carbon atom | $\mathrm{AC} 6 \mathrm{H} \cdot \mathrm{Na}$ Chemical shift $\delta$ (ppm) | AC6D•Na Chemical shift $\delta$ (ppm) | Carbon atom | AC6H•Na Chemical shift $\delta$ (ppm) | AC6D•Na Chemical shift $\delta(\mathrm{ppm})$ |
| :---: | :---: | :---: | :---: | :---: | :---: |
| 1 | 175.1 | 175.1 | $1^{\prime}$ | 97.3 | 96.8 |
| 2 | 98.5 | 98.5 | $2^{\prime}$ | 39.7 | 35.8 |
| 3 | 201.0 | 200.9 | $3^{\prime}$ | 53.1 | 91.5 |
| 4 | 52.0 | 52.0 | $4{ }^{\prime}$ | 58.1 | 54.0 |
| 5 | 39.2 | 39.1 | 5 | 69.8 | 69.1 |
| 6 | 31.4 | 31.4 | 6 | 17.3 | 17.0 |
| 7 | 42.3 | 42.3 | $7{ }^{\prime}$ | 28.6 | 25.4 |
| 8 | 35.0 | 35.0 | $8{ }^{\prime}$ | 158.4 | 158.4 |
| 9 | 75.3 | 75.3 | $9{ }^{\prime}$ | 52.5 | 52.8 |
| 10 | 44.5 | 44.4 | * | 171.2 | 171.2 |
| 11 | 128.1 | 128.0 | * | 99.6 | 99.6 |
| 12 | 125.7 | 125.8 | * | 98.8 | 98.8 |
| 13 | 51.7 | 51.8 | * | 92.8 | 92.8 |
| 14 | 138.1 | 138.1 | * | 92.1 | 92.1 |
| 15 | 121.9 | 121.7 | * | 85.6 | 85.6 |
| 16 | 31.2 | 31.2 | * | 81.4 | 81.4 |
| 17 | 78.9 | 78.9 | * | 74.9 | 75.0 |
| 18 | 138.6 | 138.7 | * | 71.4 | 71.4 |
| 19 | 121.1 | 120.7 | * | 70.5 | 70.5 |
| 20 | 45.4 | 45.5 | ** | 68.2 | 68.2 |
| 21 | 70.5 | 70.5 | * | 67.0 | 67.0 |
| 22 | 152.2 | 151.1 | * | 63.5 | 63.5 |
| 23 | 137.0 | 137.3 | * | 62.3 | 62.3 |
| 24 | 29.7 | 29.7 | * | 37.7 | 37.8 |
| 25 | 83.9 | 83.9 | * | 31.7 | 31.6 |
| 26 | 200.2 | 200.1 | ** | 29.7 | 29.7 |
| 27 | 15.1 | 15.1 | * | 27.2 | 27.3 |
| 28 | 22.4 | 22.4 | * | 26.5 | 26.5 |
| 29 | 14.3 | 14.3 | * | 21.0 | 21.0 |
| 30 | 14.5 | 14.6 | * | 18.8 | 18.8 |
| 31 | 15.8 | 15.8 | * | 18.0 | 18.0 |
| 32 | 194.3 | 194.4 | * | 17.8 | 17.8 |
|  |  |  | * | 17.6 | 17.6 |

Asterisks $\left({ }^{*, * *}\right)$ were correlated with the carbons belonging to amicetose and digitoxose moieties and were not fully assigned yet.
Asterisks ( ${ }^{* *}$ ) should the presence of two carbon signals.

Fig. 1. ${ }^{13} \mathrm{C}$ NMR spectra of $\mathrm{AC} 6 \mathrm{H} \cdot \mathrm{Na}$ and $\mathrm{AC} 6 \mathrm{D} \cdot \mathrm{Na}\left(270 \mathrm{MHz}, \mathrm{CDCl}_{3}-\mathrm{CD}_{3} \mathrm{OD}\right)$.

was concentrated to a small volume ( 350 ml ) in vacuo, and it was extracted with chloroform ( $100 \mathrm{ml} \times 3$ ) after dilution with water ( 150 ml ). All chloroform extracts were combined and dried in vacuo to afford a crude AC6 complex, that was further purified by silica gel chromatography using $\mathrm{CHCl}_{3}-\mathrm{MeOH}$ $\mathrm{H}_{2} \mathrm{O}(85: 15: 1.5 \mathrm{v} / \mathrm{v})$ as a solvent system to give two active fractions, designated as crude AC6D and AC6H fractions. The AC6H fraction was concentrated and applied to preparative silica gel TLC using the same solvent system and the band of Rf 0.29 corresponding to $\mathrm{AC} 6 \mathrm{H} \cdot \mathrm{Na}$ was extracted with MeOH to give a pale yellow powder ( 29.7 mg ). It showed a single HPLC peak at 18.4 minutes by a Senshu Pak ODS-3251-H column ( $8 \times 250 \mathrm{~mm}$, solvent $\mathrm{MeOH}-\mathrm{H}_{2} \mathrm{O}(7: 3)$, flow rate $1.5 \mathrm{ml} /$ minute $)$ monitored with UV absorption at 254 nm . The AC6D fraction gave a component of AC6D•Na ( $84 \mathrm{mg}, \mathrm{mp} 236 \sim 241^{\circ} \mathrm{C}$ ) by the same procedures. AC6D $\cdot \mathrm{Na}$ was treated with dil. HCl to afford a free AC6D compound. The spectral data and the physico-chemical properties of AC6D (UV, IR, NMR and mass) are in good agreement with the data published for tetrocarcin $\mathrm{A}^{1 \sim 5)}$ and antlermi$\operatorname{cin} A^{6}$.
Antibiotic AC6H, slightly yellow amorphous powder of mp $239 \sim 241.5^{\circ} \mathrm{C}$ (dec), has a molecular formula of $\mathrm{C}_{67} \mathrm{H}_{98} \mathrm{O}_{22} \mathrm{~N}_{2}$ (HRFAB-MS: Calcd: $m / z$ 1283.6689, Found $\left.m / z 1283.6680(\mathrm{M}+\mathrm{H})^{+}\right),[\alpha]_{\mathrm{D}}$ (MeOH) -82.64 , absorption peaks at $\lambda_{\text {max }}^{\mathrm{MeOH}} \mathrm{nm}(\varepsilon)$ $235(19,660), 266(13,130), 278(9970 \mathrm{sh})$ in UV region and $v_{\text {max }}(\mathrm{KBr}) 3400,2900,1730,1710,1640$, $1620,1530,1240,1120,1050 \mathrm{~cm}^{-1}$ in IR region. ${ }^{13} \mathrm{C}$ NMR spectrum of AC6H was closely related
with that of tetrocarcin A (AC6D) as shown in Table 1. Precise comparison of these spectra (Fig. 1) revealed the same chemical shifts for 59 out of 63 carbons of AC6H and tetrocarcin A (AC6D), and the remaining 4 carbons distributed around $\mathrm{C} 17 \sim 3^{\prime}$ of tetrocarcin A . The quarternary $\mathrm{C}-\mathrm{NO}_{2}$ signal ( $\delta$ $91.5 \mathrm{ppm})$ of tetrocarcin A was replaced by the quarternary carbon ( $\delta 53.1$ ) of AC6H. AC 6 H has a molecular weight of 1283,30 mass units less than that of tetrocarcin A.

Mild acid hydrolysis of AC 6 H with 0.2 N hydrogen chloride in acetone at $60^{\circ} \mathrm{C}$ for 1 hour gave a tetronolide derivative, which had the same difference in molecular weight with the hydrolysate of tetrocarcin A, tetronitrosyl tetronolide, and strong acid hydrolysis of AC 6 H at $80^{\circ} \mathrm{C}$ for 2 hours gave the same aglycone as tetrocarcin A and an additional sugar component, designated as Asugar. The structure of A-sugar was determined by the analyses of mass and NMR spectra. The EI mass spectrum of TMS-A-sugar revealed no molecular ions, but the ions at $m / z 115,130,158$, 176, 214 and 231 were diagnostic for the structure of the sugar compound. The FAB-MS spectrum of A-sugar gave a $(\mathrm{M}+\mathrm{Na})^{+}$ion at $m / z 241$. ${ }^{13} \mathrm{C}$ NMR data were in agreement with the data reported by Alan K. Mallams ${ }^{7}$ ) for $2,3,4,6$-tetra-deoxy-4-(methoxy-carbonylamino)-3-C-methyl-3-amino-D-xylo-hexopyranosyl structure, which was prepared by the reduction of tetronitrose, a sugar moiety of tetrocarcin A, with Raney nickel. These results clearly suggested that the structural difference of AC6H with AC6D existed at the nitrosugar moiety of tetrocarcin A, and AC6H had a $\mathrm{C}-\mathrm{NH}_{2}$

Fig. 2. Structures of AC 6 H and AC 6 D .


Table 2. Antitumor activity of AC 6 D and AC 6 H on P388 leukemia.

| Sample | Dose <br> $(\mathrm{mg} / \mathrm{kg})$ | Survival <br> days | $\mathrm{T} / \mathrm{C}$ <br> $(\%)$ |
| :--- | :---: | :---: | :---: |
| AC6H | 100 day $1,5,9$ | Toxic |  |
|  | 50 day $1,5,9$ | 11.8 | 118 |
|  | 25 day $1,5,9$ | 11.0 | 110 |
| AC6D | 100 day $1,5,9$ | 15.0 | 150 |
| Adriamycin | 5 day 1 | $>15$ | $>150$ |
| Control |  | 10.0 | 100 |

group instead of the $\mathrm{C}-\mathrm{NO}_{2}$ group of tetrocarcin A (AC6D) as shown by the ninhydrin positive character (Fig. 2).
Antitumor activity of AC6H was examined using P388 leukemia cells (CDF1 mouse, 6 weeks) to show slight elongation of mean survival days (Table 2). Cytotoxicity of AC6H against P388 leukemia and B16 melanoma cells were 6.25 and $25 \mu \mathrm{~g} / \mathrm{ml}$ respectively, whereas those of AC6D (tetrocarcin A) were 0.39 and $0.78 \mu \mathrm{~g} / \mathrm{ml}$.
In conclusion, we have elucidated the structure of a new antitumor antibiotic, AC6H by spectroscopic comparison with tetrocarcin A and chemical derivation from tetrocarcin A .

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